e-ISSN: 2792-4017 | www.openaccessjournals.eu | Volume: 1 Issue: 4

## Preliminary Study on GC-MS Analysis of Prosopis Africana Seed (African Mesquite) Oil

#### Agubosi, O. C. P.; Oluwafemi, R. A; Alagbe, J. O

Department of Animal Science, University of Abuja, Gwagwalada, Nigeria

**ABSTRACT:** A preliminary studywas carried out to examine the various bioactive compounds in *Prosopisafricana* seed (*African mesquite*)oil usinggas chromatography-mass spectrometry (GC/MS). A total number of 24 bioactive compounds was identified including: 2-phenyl propane (3.09 %), 1,3,5-trimethyl benzene (4.80 %), prosogerin A (12.67 %), prosogerin B (10.06 %), prosogerin C (7.11 %), prosogerin D (4.05 %), prosogerin E (2.89 %), luteolin 7-Oβ-D-glucoside (0.67 %), quercetin 3-O-rutinoside (4.88 %), isoprosopilosine (4.28 %), tryptamine (1.66 %), βphenethylamine (5.63 %), gallic acid (0.57 %), 4-hydroxy benzoic acid (1.41 %), 2,4-bis (1-phenylethyl) phenol (9.60 %), 2,4,6-tris(1-phenylethyl) phenol (7.83 %), rhodoxanthin (1.67 %), ferulic acid (1.20 %), pheophytin A (1.54 %), pheophytin B (3.31 %), campsterol (2.80 %) and β-sitosterol (1.51 %).Prosogerins are the most abundant compounds in the sample and are known to posses antimicrobial, anti-inflammatory and capable of scavenging free radicals thus preventing diseases in animals. It was concluded that *Prosopisafricana* seed oil can be used as potential alternative to antibiotics and can be used to bridge the gap between food safety and livestock production.

Key words: Prosopisafricana, seed, oil, phytochemicals, free radicals

#### Introduction

Seeds from plants are potential reservoirs for bioactive compounds or secondary metabolites, proteins, carbohydrates, amino acid and fats. Recently, there has been increasing awareness on food safety and the use of plants as potential alternatives to antibiotics due to antimicrobial resistance, toxic residues of synthetic drugs in livestock products, increase in different ailments and death (Singh *et al.*, 2021). Seeds from plants of medicinal origin contains essential oils, which have been proven to be rich in phytochemicals (tannins, flavonoids, phenols, saponins, alkaloids, terpenoidsetc), effective and generally regarded as safe (Adewale*et al.*, 2021). The high cost of animal protein has directed interest towards several seed protein as potential sources of vegetable protein for human food (Musbau and Asiru, 2020).

*Prosopisafricana* (*African mesquite*) is a tropical leguminous plant belonging to the family Fabaceae. The tree reaches about 4-20 m in height; has an open crown and slightly rounded buttresses; bark is very dark, scaly, slash, orange to red-brown with white streaks (Aremu*et al.*, 2007). Seeds from *Prosopisafricana*are used to prepare traditional fermented soup condiment or as flavor enhancers in Northern parts of Nigeria (Tajudeen*et al.*, 2011). The seeds are rich in several bioactive compounds which makes a medicinal plant (Oloyede, 2005). The ripe pods of *P. africana*are harvested by shaking off the ripe pod from the tree branches. According to Musbau and Asiru (2020) proximate composition of fermented *P. africana* seeds and pods revealed the presence of crude protein (27.75 %, 7.62 %), carbohydrate (27.09 %, 72.72 %), ether extracts (10.52 %, 6.86 %), fibre (6.76 %, 3.28 %), moisture (0.03 %, 5.62 %) and ash (6.04 %, 3.90 %). Oluwafemi*et al.* (2021) also reported that *P. africana* seed oil contains crude protein, crude fibre, ether extract, ash and energy at 30.71 %, 6.47 %, 3.66 %, 5.08 % and 383.26 (Kcal/kg).

The pods are useful in the feeding of ruminant animals due to their high nutritive value and can also be used traditionally for the treatment of tooth ache and other skin infections (Olorunmaiye*et al.*, 2019).

This experiment was designed to examine the secondary metabolites in *Prosopisafricana* seed oil using GC-MS. It will further give a clue on its medicinal properties as well as the physiological actions of the oil in the body of animals.

#### Materials and methods

#### Experimental site

This study was carried out at the department of Animal Science Research Laboratory, University of Abuja, along airport road, Gwagwalada, Abuja, Nigeria.Gwagwalada is located between latitudes  $8^{\circ}57^{1}$  and  $8^{\circ}55^{1}$ N and longitude  $7^{\circ}05^{1}$  and  $7^{\circ}06^{1}$ E.

### e-ISSN: 2792-4017 | www.openaccessjournals.eu | Volume: 1 Issue: 4

#### Extraction of Prosopisafricana seed

Mature seeds from *Prosopisafricana* were harvested from different trees at the University of Abuja, Teaching and Research Farm. It was identified and authenticated by a certified taxonomist at the Department of Biological Sciences, University of Abuja, Gwagwalada with a voucher specimen number ANS/08F/2020. The seeds were sorted to remove the bad ones and washed with running water to remove dirt's and air dried for 18 days to maintain the bioactive chemicals in the seeds, mechanically separated from their pods with knife to obtain the seeds. Dried seeds was grinded using a laboratory grinder (Panasonic: model AS/309F) to obtain *Prosopisafricana* seed meal and stored in a clean well labeled container for analysis. Prior to the commencement of the analysis, laboratory equipmentswere serviced and all the necessary reagents were purchased.

Extraction of PASO was done using a cold press machine (Model: YZYX168, China). The grinded *Prosopisafricana* (2000 g) was poured at into the feeder of the machine which works at a low temperature and PASO was collected via the squeeze cage.

#### Gas chromatography – Mass spectrometry analysis (GC-MS)

GC-MS analysis were performed on a GC – 2010 Shimadzu capillary gas chromatography directly coupled to the mass spectrometer system (GC-MS – model QP 2010; S/N column (70464300019 SA; Shimadzu) DB – 5ms non polar fused silica capillary column (30m X 0.25mm, 0.25 $\mu$ m film thickness) was used under following conditions: oven temperature program isotherm 2 min at 70°C, 3°C/min gradient to 200°C and final temperature kept for 35 min; injection temperature 200°C carrier gas is helium with flow rate 1.51ml/min; linear velocity 45.1 cm/sec. The effluent of the GC column was introduced directly into the source of MS and spectra obtained in the EI mode with ionization energy 70eV, in the electronic ionization mode and ion source temperature is 200°C. The solvent cut time 3 min. The sector mass analyzer was set to scan from 40 to 1000m/z with interface temperature of 240°C. The components of the essential oil were identified on the basis of comparison of their relative indices and mass spectra by computer matching with National Institute of Standards and Technology (NIST08) libraries.

#### Gas chromatography mass spectrophotometer (GC-MS) composition of Prosopisafricana oil

The GC-MS result of Prosopisafricana seed oil revealed many bioactive compounds. These compounds with their molecular weight, molecular formulae, retention time and their composition (%) determined from their peak areas is presented in Table 1. A total number of 24 bioactive compounds was identified including: 2-phenyl propane (3.09 %), 1,3,5-trimethyl benzene (4.80 %), prosogerin A (12.67 %), prosogerin B (10.06 %), prosogerin C (7.11 %), prosogerin D (4.05 %), prosogerin E (2.89 %), luteolin 7-O-β-D-glucoside (0.67 %), quercetin 3-O-rutinoside (4.88 %), isoprosopilosine (4.28 %), tryptamine (1.66 %),  $\beta$ -phenethylamine (5.63 %), gallic acid (0.57 %), 4-hydroxy benzoic acid (1.41 %), 2,4-bis (1-phenylethyl) phenol (9.60 %), 2,4,6-tris(1-phenylethyl) phenol (7.83 %), rhodoxanthin (1.67 %), ferulic acid (1.20 %), pheophytin A (1.54 %), pheophytin B (3.31 %), campsterol (2.80 %) and β-sitosterol (1.51 %). The result obtained in this study agrees with the report of Peter et al. (2009); Ferguson et al. (2005); Bhardwaj et al. (1979, 1981); Simpson and Solbrig (1977). Bioactive compounds in plants are also regarded as phytochemicals which have been proven to be relatively cheap, safe effective and recently projected as suitable alternative to antibiotics (Adewaleet al., 2021; Musa et al., 2020). For instance, prosogerins are group of flavonoids which performs multiple pharmacological functions such as: antibacterial, anti-inflammatory, antiviral, antitumor, hypolipidaemic, antiprotozoal, cytoprotective, antioxidant, antiplatelet and so on (Harzallahand Jannet, 2005; Alagbe and Motunrade, 2019; Alagbe, 2019). Prosogerins (A, B, C, D and E) are the most abundant compounds in this study; this confirms the earlier findings of Valliet al. (2014). Isoprosopilosine, tryptamine and  $\beta$ -phenethylamine are group of alkaloids which have diverse physiological effects: antibacterial, antimitotic, anti-inflammatory, analgesic, local anesthetic, hypnotic, psychotropic, and antitumor activity and many others (Aneelaet al., 2014; Ukaniat al., 2000). Alkaloids are also capable of working as an antimicrobial, antispasmodic and scavenging free radicals to prevent disease in the body of animals (Alagbe and Motunrade, 2019; Latif et al., 2003). Studies have also reported the beneficial effects of phenol as an antioxidant, therefore the presence of 2,4-bis (1-phenylethyl) phenol and 2,4,6-tris(1-phenylethyl) phenols is a clear indication that Prosopisafricana seed oil can function as an antibacterial (Kasoloet al., 2010; Singh et al., 2021) and can be traditionally used to treat skin diseases, wounds and other ailments.

Various bioactive chemicals in *Prosopisafricana* seed oil exhibit a wide spectrum of antibacterial activities against pathogenic organisms thus influencing voluntary intake in animals and bile secretion (Zeng *et al.*, 2015). Composition of *Prosopisafricana* seed oil varies according to plant species, geographical origin, season, environmental factors, processing techniques and storage conditions (Gadde*et al.*, 2017).

## e-ISSN: 2792-4017 | www.openaccessjournals.eu | Volume: 1 Issue: 4

Table 1 Major chemical compounds of *Prosopisafricana* oil as detected by gas chromatography mass spectrophotometer (GC-MS)

Chemical compounds	RI	% Area	MW(g/mol)	MF	Mass peak
2-phenyl propane	796	3.09	136.2	$\underline{C}_{9}\underline{H}_{12}\underline{O}$	27, 42, 54, 79
1,3,5-trimethyl benzene	951	4.80	120.2	C <sub>9</sub> H <sub>12</sub>	27, 55, 101, 134
Prosogerin A	1224	12.67	312.3	$\underline{C}_{17}\underline{H}_{12}\underline{O}_{6}$	27, 87, 102, 141
Prosogerin B	1009	10.06	330.8	$C_{18}H_{14}O_7$	27, 45, 58, 105
Prosogerin C	806	7.11	372.4	$\underline{C}_{20}\underline{H}_{20}\underline{O}_7$	27, 62, 71, 90, 166
Prosogerin D	1205	4.05	358.3	$\underline{C}_{19}\underline{H}_{18}\underline{O}_7$	27, 41, 48, 54, 71
Prosogerin E	1104	2.89	344.3	C18H16O <sub>7</sub>	27, 38, 51, 74, 89
Luteolin 7-O-β-D-glucoside	1967	0.67	447.4	$C_{21}H_{19}O_{11}$	27, 38, 77, 96, 155
Quercetin 3-O-rutinoside	2196	4.88	610.51	C <sub>27</sub> H <sub>30</sub> O <sub>16</sub>	27, 63, 89, 94, 106
Apigenin-8-glucoside	1663	3.31s	564.5	C <sub>26</sub> H <sub>28</sub> O <sub>14</sub>	27, 47, 69, 76, 93
Quercetin-3-glucoside	1907	5.43	464.09	$C_{21}H_{20}O_{12}$	27, 85, 89, 94, 97
Isoprosopilosine	1800	4.28	933.4	$\underline{C}_{57}\underline{H}_{104}\underline{O}_9$	27, 41, 55, 81, 95
Tryptamine	2506	1.66	160.2	$C_{10}H_{12}N_2$	27, 41, 57, 93. 121
β-phenethylamine	1002	5.63	121.18	$C_8H_{11}N$	27, 67, 80, 89, 93
Gallic acid	2707	0.57	170.12	$C_7H_6O_5$	27, 88, 89, 97, 109
4-hydroxy benzoic acid	779	1.41	138.12	$C_7H_6O_3$	27, 40, 43, 86, 92
2,4-bis (1-phenylethyl) phenol	1117	9.60	330.5	$C_{24}H_{26}O$	27, 45, 57, 66, 72
2,4,6-tris(1-phenylethyl) phenol	1251	7.83	406.6	$C_{30}H_{30}O$	27, 48, 53, 67
Rhodoxanthin	2265	1.67	562.9	$C_{40}H_{50}O_2$	27, 40, 43, 48, 60
Ferulic acid	1307	1.20	194.18	$C_{10}H_{10}O_4$	27, 38, 40, 54
Pheophytin A	922	1.54	871.2	$C_{55}H_{74}N_4O_5$	27, 42, 47, 76, 78
Pheophytin B	1006	3.31	885.2	$C_{55}H_{72}N_4O_6$	27, 34, 37, 38
Campsterol	1227	2.80	400.7	C <sub>28</sub> H <sub>48</sub> O	27, 32, 43, 47
β-sitosterol	2093	1.51	414.71	$C_{29}H_{50}O$	27, 45, 30, 45, 52

RI: retention index; MW: molecular weight; MF: molecular formula

#### Conclusion

*Prosopisafricana* seed oil contains chemical compounds with recognized medicinal value/therapeutics, or which are precursors for chemo-pharmaceutical semi-synthesis. The medicinal value of this oil lies in some chemical substances (phytochemicals or bioactive chemicals) that produce a definite physiological action in the body of animals. The major bioactive compounds in *Prosopisafricana* seed oil are prosogerins and their compositionand concentration vary according to the plant, parts of the plant, geographical origin, harvesting season, environmental factors, storage conditions, and processing techniques.

#### References

- 1. Kasolo, J.N., Gabriel, S., Bimenya, L.O., Joseph, O. and Ogwal-Okeng, J.W. (2010). Phytochemicals and uses of Moringaoleifera leaves in Ugandan rural communities. J. Med. Plants Res. 4(9): 753-757.
- 2. Latif A, Ahmad H, Begum S, Adnan M, Hassian S, Waseem M. (2003). Medicinal and other economic plants as substitute to forest logging in Miandam and Sulatanr valleys, Swat. Proceedings of international workshop on conservation and sustainable use of medicinal and aromatic plants in Pakistan. WWF Pak., 101-105
- 3. Gadde, U., Kim, W.H., Oh, S.T and Lillehoj, H.S. (2017). Alternatives to antibiotics for maximizing growth and feed efficiency in Poultry: A review. Journal of Animal Science Research. 18: 26-45.
- 4. **Musa, B.,**Alagbe, J.O.,AdegbiteMotunrade Betty, Omokore, E.A. (2020).Growth performance, caeca microbial population and immune response of broiler chicks fed aqueous extract of *Balanitesaegyptiaca* and *Alchorneacordifolia* stem bark mixture. *United Journal for Research and Technology*, 2(2):13-21.
- 5. Alagbe, J.O and AdegbiteMotunrade Betty (2019). Haematological and serum biochemical indices of starter broiler chicks fed aqueous extract of *Balanitesaegyptiaca* and *Alchorneacordifolia* bark mixture. *International Journal of Biological, Physical and Chemical Studies*. 1(1): 8-15
- 6. Alagbe, J.O (2020). Proximate, phytochemical and vitamin compositions of *Prosopisafricana*stem bark. *European Journal of Agricultural and Rural Education*. 1(4): 1-7.

### e-ISSN: 2792-4017 | www.openaccessjournals.eu | Volume: 1 Issue: 4

- 7. Oluwafemi, R.A., Akinbisola, S.A and Alagbe, J.O. (2020). Nutritional and growth performance of feeding *Polylathialongifolia* Leaf Meal as partial replacement of Wheat Offal in the diet of broiler chicks. *European Journal of Biotechnology and Bioscience*. 8(4): 17-21.
- 8. Singh, A.S., Alagbe, J.O., Sharma, S., Oluwafemi, R.A and Agubosi, O.C.P. (2021). Effect of dietary supplementation of melon (*Citralluslinatus*) seed oil on the growth performance and antioxidant status of growing rabbits. *Journal of Multidimensional Research and Reviews*, 2(1): 78-95.
- 9. Alagbe, J.O. (2021). *Prosopisafricana* stem bark as an alternative to antibiotic feed additives in broiler chicks diets: Performance and Carcass characteristics. *Journal of Multidimensional Research and Reviews*, 2(1): 64-77.
- 10. Adewale, A.O., Alagbe, J.O., Adeoye, Adekemi. O. (2021). Dietary Supplementation of *RauvolfiaVomitoria* Root Extract as A Phytogenic Feed Additive in Growing Rabbit Diets: Haematology and serum biochemical indices. *International Journal of Orange Technologies*, 3(3): 1-12.
- 11. Oloyede, I.O. (2005). Chemical profile of Unripe pulp of Carica papaya. Pakistan Journal of Nutrition 6: 379-381.
- 12. Tajudeen, A.I., Kayode, C.O., Yahaya, A and Mohammed, S.A. (2011). Development and testing of a Prosopis pod thresher. Australian Journal of Basic and Applied Sciences 5(2): 759-767.
- 13. Olorumaiye, K.S., Apeh, L.E., Madandola, H.A and Oguntoye, M.O. (2019). Proximate and phytochemical composition of African mahogany seed and *African mesquite* pod. Journal of Applied Sciences and Environmental Management 23(2): 249-252.
- 14. Musbau, S and Asiru, R.A. (2020). Proximate parameters of fermented Prosopisafricana seeds. Journal of Academia and Industrial Research 8(9): 163-165.
- 15. Aremu, M.O., Olanisakin, B.O., Atolaye, B.O and Ogbu, C.F. (2007). Some nutritional composition and functional properties of *Prosopisafricana*. Bangladesh Journal of Industrial Research 4(3): 269-270.
- 16. Aneela, S., Dey, A and De, S. (2014). GC-MS analysis of methanolic extract of *Prosopisspicigera*. International Journal of Phytopharmacology 5: 168-171.
- 17. Ukani, M.D., Limbani, N.B and Mehta, N.K. (2000). A review on the Ayurvedic herb *Prosopis cineraria* (L) Druce. Anc. Sci. Life 20:58-70
- 18. Simpson, B.B and Solbrig, O.T. (1977). Introduction Mesquite: its biology in two desert ecosystems. Inc. Stroudsberg, pp 1-7.
- 19. Peter, S., Stephene, D., Eric, M and Phillipe, G. (2009). Unusal amount of (-) mesquitol from the heartwood of *Prosopisjuliflora*. Nat. Prod. Res. 23: 183-189.
- Valli, S., Gokulshankar, S., Mohanthy, B.K., Ranjith, M.S., Ashutosh, S.R and Remya, V. (2014). Anticrytococcal activity of alkaloid rich fraction of leaves of *Prosopisjuliflora*. A future promising supplementary therapy for cryptococcosis and cyyptococcal meningitis. Int. J. Pharm. Sci. 6: 491-495.
- Harzallah, S.F and Jannet, H.B. (2005). Flavonoids diversification in organs of two Prosopisfracta (Bank and Sol.) eig. (Leguminosea, Mimosoidae) populations occurring in Northeast and the Southeast of Tunisia. Journal of Applied Research 1: 130-136.
- 22. Ferguson, L.R., Shuo-tun, Z and Harris, P. (2005). Antioxidant and antigenotoxic effects of plant cell wall hydroxycinnamic acids in cultured HT-29 cells. Molecular Nutrition and Food Research 49: 585-593.
- 23. Bhardwaj, D.K., Bisht, M.S., Mehta, C.K and Sharma, G.C. (1979). Flavonoids of *Prosopisspicigera* flowers. Journal of Phytochemistry 18: 355-356.
- 24. Bhardwaj, D.K., Gupta, A.K., Jain, R.K and Sharma, G.C. (1981). Chemical examination of *Prosopisspicigera* seeds. J. Nat. Prod. 44: 656-659.
- 25. Oluwafemi, R.A., Agubosi, O.C.P and Alagbe, J.O. (2021). Proximate, minerals, vitamins and amino acid composition of prosopisafricana (*African mesquite*)seed oil. Asian Journal of Advances in Research 11(1): 21-25.